

The immune system and autism spectrum disorder: association and therapeutic challenges

Arash Heidari^{1,2}, Yasna Rostam-Abadi^{3,4} and Nima Rezaei^{2,4,5*}

¹ School of Medicine, Tehran University of Medical Sciences, Tehran, Iran,

² Network of Immunity in Infection, Malignancy and Autoimmunity (NIIMA), Universal Scientific Education and Research Network (USERN), Tehran, Iran,

³ Iranian National Center for Addiction Studies, Tehran University of Medical Sciences, Tehran, Iran,

⁴ Research Center for Immunodeficiencies, Children's Medical Center, Tehran University of Medical Sciences, Tehran, Iran,

⁵ Department of Immunology, School of Medicine, Tehran University of Medical Sciences, Tehran, Iran,

*Email: rezaei_nima@tums.ac.ir

Autism spectrum disorder (ASD) is a neurodevelopmental disorder, affecting communication and behavior. Historically, ASD had been described as a purely psychiatric disorder with genetic factors playing the most critical role. Recently, a growing body of literature has been emphasizing the importance of environmental and immunological factors in its pathogenesis, with the autoimmune process attracting the most attention. This study provides a review of the autoimmune involvement in the pathogenesis of ASD. The microbiome, the representative of the innate immune system in the central nervous system (CNS), plays a critical role in triggering inflammation. Besides, a bidirectional communicational pathway between the CNS and the intestine called the gut-brain-axis is linked to the development of ASD. Moreover, the higher plasma level of pro-inflammatory cytokines in ASD patients and the higher prevalence of autoimmune disorders in the first-degree family members of affected persons are other clues of the immune system involvement in the pathogenesis of ASD. Furthermore, some anti-inflammatory drugs, including resveratrol and palmitoylethanolamide have shown promising effects by relieving the manifestations of ASD. Although considerable advances have been made in elucidating the role of autoimmunity in the ASD pathogenesis, further studies with stronger methodologies are needed to apply the knowledge to the definitive treatment of ASD.

Key words: autism spectrum disorder, autoimmune diseases, inflammation

INTRODUCTION

Autism spectrum disorder (ASD) is a neurodevelopmental disorder, characterized by persistent impairment in social communication and restricted, repetitive, and stereotypical patterns of interests, behaviors, and activities (American Psychiatric Association, 2013). The word “spectrum” implies a range of heterogeneous symptoms with various severities and difficulty levels, providing a unique clinical definition for associated disorders (Paglia, 2020). According to the Diagnostic and Statistical Manual of Mental

Disorders-V (DSM-V), the previously named mental disorders, including Asperger's disorder, autistic disorder, the pervasive developmental disorder not otherwise specified, and childhood disintegrative disorder, are now described under the umbrella term of ASD (American Psychiatric Association, 2013; Paglia, 2020).

Epidemiological studies have shown discrepancies in the estimation of the prevalence of ASD. For instance, the estimated prevalence for autistic disorder in 36 studies investigated by Fombonne (2005) ranged from 0.7 to 72.6 in 10,000. These variable estimates can be attributed to different factors, including inconsis-

tent definitions of the disease and different reporting practices (Hansen et al., 2015). However, most studies proposed that the prevalence of ASD tends to increase over the past 50 years (Fombonne, 2005). In its updated guidelines in 2016, the United States' Center for Disease Control and Prevention reported the average incidence of ASD among US children aged eight years to be about 1.46% (Bjorklund et al., 2020). Besides, ASD has been reported to occur in all racial, ethnic, and socioeconomic groups (Baio, 2014). A meta-analysis study, conducted by Loomes et al. (2017) showed males are three times more susceptible to develop ASD, compared to females. These different prevalence rates can be explained by the fact that girls with ASD are less willing to receive the clinical diagnosis because of the possibility to mask social deficits in the female population, using a process called “camouflaging” (Volkmar et al., 2014; Loomes et al., 2017).

Although ASD has been extensively investigated during recent decades, etiological aspects of the disease have remained relatively unclear. Previous studies suggested that the interaction of genetic and environmental factors with the immune system could explain the condition (Gottfried et al., 2015). At the same time, researchers proposed some risk factors for ASD, including immunological abnormalities, advanced parental age, maternal treatment with some pharmaceutical drugs such as selective serotonin reuptake inhibitors (Matelski and Van de Water, 2016), preterm labor (Chung et al., 2020), male sex (Baio, 2014), and the presence of some inherited abnormalities like fragile X syndrome (Kaufmann et al., 2017). Among these risk factors, immunological factors have attracted so much attention so that a significant body of literature is forming, pointing to the role of the immune system as a possible etiology of ASD (Sciara et al., 2020). Furthermore, several immunological conditions, including gestational immune abnormalities, family history of autoimmunity, and coexistence of autoimmune diseases in individuals with ASD have been proposed to play a role in developing this disease (Edmiston et al., 2017). Anatomical studies revealed evidence of some brain structural changes related to neuroinflammation in patients with ASD. Epidemiological studies have also shown some evidence of simultaneously increased risk of ASD with the outbreak of atopic diseases in some societies (Liao et al., 2016; Sciara et al., 2020). Moreover, the presence of inflammatory cytokines in some patients with autism has demonstrated another clue of the immune system involvement in the pathological mechanism underlying ASD (Saghazadeh et al., 2019b). Accordingly, this paper aims to review the role of the immune system in the pathogenesis of ASD.

The interaction of the immune and nervous systems

The immune and nervous systems are among the most complicated body systems whose functions have some similarities. For example, both systems transport and receive messages *via* the secretion of transmitter molecules. They are also involved in storing data, creating the memory, and retrieving information (Habibi et al., 2009). Besides, both are capable of suppressing or strengthening their responses. The T/B regulatory cells in the immune system play a crucial role in suppressing immune responses for self-antigens (Sakaguchi et al., 2008; Rosser and Mauri, 2015). The inhibitory and excitatory subdivisions of neurons balance the neural system to function correctly (Yizhar et al., 2011).

The nervous system, divided into the central and peripheral systems, can influence the immune system either by straight innervation of immune organs or by secreting immune-modulatory hormones *via* the hypothalamic-pituitary-adrenal axis (Dunn, 2001; Nance and Sanders, 2007). The central nervous system (CNS), including the brain and the spinal cord, comprises neurons and glial cells. The glial cells in the mature CNS consist of oligodendrocytes, astrocytes, and microglia (Purves et al., 2001), each involved in secreting cytokines and triggering an immune response in the CNS. However, the microglial cells are the most prominent contributors in this regard (Bitzer-Quintero and González-Burgos, 2012).

The role of microglia in the pathogenesis of ASD

Microglial cells, characterized by their healing functions in local brain injuries (Streit, 1996), are macrophage-derived cells residing in the CNS. They are in charge of secreting different immune mediators, including pro-inflammatory and anti-inflammatory cytokines, prostanoids, chemokines, and thromboxane-A₂ (Bitzer-Quintero and González-Burgos, 2012). Moreover, they have an essential role not only in the process of neurogenesis but also in neural death (Kettenmann et al., 2011). They are located in the brain parenchymal or perivascular regions and play a significant role in representing the innate immune system in these areas. When activated as antigen-presenting cells (APC), they trigger the secretion of T helper-1 (TH1)-derived cytokines, including IL-7 and tumor necrosis factor-alpha (TNF-alpha) (Bitzer-Quintero and González-Burgos, 2012). With the secretion of such immune mediators, the perivascular microglia signal and activate the parenchymal microglia,

which release different pro-inflammatory cytokines, resulting in CNS inflammation (Bitzer-Quintero and González-Burgos, 2012). Moreover, microglia affects the neurons in both embryonic and adult life, either indirectly *via* cytokines or by direct contact (Bilimoria and Stevens, 2015).

Recent studies have demonstrated that microglial functions are vital for brain development. They do this by taking the patterning and wiring processes of the developing CNS under control through programmed cell death, synaptic pruning, and synaptic maturation (Bilimoria and Stevens, 2015). Synaptic pruning is the process of the elimination of the extra non-required synapses in the development of the CNS. Several neurodevelopmental disorders, including socio-behavioral defects like ASD, have been linked to failure in this process (Kim et al., 2017). Furthermore, the C1q and C3, parts of the complement's cascade, are other contributors in the synaptic pruning process by their roles in assisting microglial functions. Therefore, impairment in these components can also be associated with neurological diseases (Stevens et al., 2007).

Role of astrocytes in the pathogenesis of ASD

Astrocytes are glial cells comprising the most extensive cell type in the brain (Freeman, 2010). Our knowledge of astrocytes was limited to mere their supporting role in CNS (Guillamón-Vivancos et al., 2015). Nevertheless, their active contribution in many CNS functions, including synapse formation, maturation, plasticity, and elimination, CNS development, construction of the BBB, neuronal plasticity, and blood flow regulation, have been discovered recently (Guillamón-Vivancos et al., 2015).

Astrocytes also play deciding roles in the pathophysiology of neurodevelopmental disorders such as ASD (Russo et al., 2018; Williams et al., 2014). Russo et al. (2018) demonstrated that astrocytes significantly impact synaptogenesis and neuronal morphological features in ASD using induced pluripotent stem cells (iPSCs). They were co-cultured *in vitro* with the normal neurons with ASD-derived astrocytes and showed morphological impairments in neurons and synaptogenesis defects (Russo et al., 2018). On the other hand, the co-culture of control-derived astrocytes with ASD-derived neurons improved ASD neuronal phenotypes and enhanced synapses revealed by higher synaptic puncta number (Russo et al., 2018). In this study, the ASD-derived neurons and the control neurons co-cultured with ASD-derived astrocytes were less complex. Furthermore, they had fewer branches than normal neurons, suggesting the pivotal role of astro-

cytes in ASD (Russo et al., 2018). Besides, the level of IL-6 was significantly higher in the ASD-derived astrocytes compared with control-derived astrocytes, and blocking IL-6 was associated with an enhancement in synaptogenesis. So it has been hypothesized that astrocyte-derived IL-6 may involve in the synaptic defect in ASD (Russo et al., 2018).

According to a post-mortal study, astrocytes are more abundant in the frontal cortex of autistic patients. Each has fewer branching processes, total branching length, and a smaller size than the astrocytes in the frontal cortex of control brains (Cao et al., 2012). Similar changes have been observed in neuroigin-3 knockdown mice used as an animal model for ASD (Cao et al., 2012). Moreover, the Wnt/ β -catenin pathway, known to be involved in astrocyte development regulation, significantly decreased in autistic brains (Cao et al., 2012).

The interaction between glial cells and neurons in ASD

Synapses are specific structures enabling a neuron to pass electrical or chemical signals to another neuron (Blanco-Suárez et al., 2017). Astrocytes in the CNS are closely associated with many synapses *via* their specialized processes called perisynaptic astrocytes' processes (PAPs) (Blanco-Suárez et al., 2017). The combination of PAPs and pre- and post-synaptic structures forms a concept named the "tripartite synapse" (Araque et al., 1999). In the developing brain, astrocytes regulate synapse formation *via* secreting varied factors, including thrombospondin, hevin, glypicans, brain-derived neurotrophic factor (BDNF), secreted protein acidic and rich in cysteine (SPARC), and TNF- α (Allen, 2014). These factors mainly induce glutamatergic synapse formation. Although not all astrocyte-derived factors lead to synapse formation, SPARC inhibits synapse formation (Kucukdereli et al., 2011). The astrocyte gliotransmitters – like ATP, adenosine, and D-serine – can act on pre- or post-synaptic compartments to strengthen or weaken neuronal transmission and plasticity (Chung et al., 2015). For example, thrombospondin inhibits presynaptic release at glutamatergic synapses (Chung et al., 2015). Excessive glutamate in synapses leads to damage and destruction of the synapse. Accordingly, astrocytes uptake the excessive glutamate in synaptic cleft *via* glutamate transporter 1 (GLT-1) and glutamate aspartate transporter (GLAST). The absorbed glutamate is then converted to glutamine in astrocytes and recycled to neurons for maintaining synaptic transmission (Blanco-Suárez et al., 2017). Furthermore, astrocytes have an active role in

buffering extracellular K(+) to support neuronal networks by rectifying potassium KIR4.1 channels (Chever et al., 2010). Given synapse abnormalities are major characteristics of neurodevelopmental disorders like ASD (Blanco-Suárez et al., 2017), changing astrocyte functions presents a novel target to improve the disease manifestations.

On the other hand, microglia and astrocytes interact with each other by secreting various cytokines and signaling molecules. For example, reactive microglia activate and proliferate astrocytes by secreting many cytokines, including IL-1, IL-2, IL-6, TNF- α , and IFN- γ (Matta et al., 2019). On the other hand, activated astrocytes discharge ATP to maintain microglial activity (Matta et al., 2019). Although these states of activity play beneficial roles in CNS injuries, the prolonged activation of these two glia can promote neuroinflammation and lead to disorders like ASD. Taken together, the extent to which the neurons and glial cells interact with each other is a crucial element for understanding the pathological mechanisms underlying various neurological diseases and leads to a better perception of the interaction of immune and nervous systems.

The innate immune system and ASD

The innate immune system is a significant contributor to the pathogenesis of autism. While this system defends the body against external pathogens, innate immunity plays a role in developing ASD by triggering inflammation in the CNS (Salam et al., 2018). Two major arms impacting the development of ASD are microglia/astrocyte and gut-brain-axis, which will be further dissected.

Microglia/astrocyte

As mentioned above, microglia and astrocytes are known as essential contributors to the pathogenesis of neurodevelopmental disorders, including ASD.

Microglia represents CNS-resident macrophages originating from the yolk sack (Gomez Perdiguero et al., 2015). They can experience different phenotypes, namely M1 and M2, to respond to different molecular and environmental stimuli and signals (Orihuela et al., 2016). Either classic or alternative way can trigger their activation. Bacterial products like lipopolysaccharide, Th1 cytokines such as IFN- γ and TNF- α , and pathogen-associated molecular patterns stimulate the classic way of activation and produce M1 phenotype (Gordon and Mantovani, 2011; Orihuela et al., 2016). M1 microglia are inflammatory cells, which secrete pro-inflammatory cytokines (such as TNF- α , IL-1 α , IL-1 β , and IL-6)

and act as microbicidal antitumorigenic agents (Zhang et al., 2016).

Moreover, these cytokines can further maintain the polarization of microglia into the M1 phenotype (Cherry et al., 2014). IL-4/IL-13 induces alternative activation and leads to M2-polarized microglia, which show anti-inflammatory features having roles in tissue repair and inflammation resolution (Orihuela et al., 2016). Depending on what cytokine stimulates the microglia activation, M2 microglia are divided into several subgroups, including M2a, M2b, and M2c. The main role of M2a is the suppression of inflammation (Cherry et al., 2014), while M2c has roles in tissue remodeling and matrix deposition (Mantovani et al., 2004), and M2b, which are less understood, can activate Th2 response and play a potential role in the initiation of M2 response (Cherry et al., 2014). Although these subgroups function somewhat differently, the common property of all of them is the production of mediators and receptors with the capacity to inhibit inflammation (Cherry et al., 2014). M1 and M2 microglia are distinguishable via arginase1+ staining (Mills, 2012).

Microglial phenotype switching is a critical concept leading to homeostasis in the CNS.

For example, in acute conditions such as traumatic brain injuries or release of DAMPs (damage-associated molecular patterns) following infections or ischemic reperfusion injury, the M1 microglia, part of the innate immune system, are activated, and secrete pro-inflammatory cytokines and reactive oxygen synthesis (ROS), leading to the elimination of invading agents and removing dead cells (Cherry et al., 2014). This inflammatory response is not only harmful, but also it is an important step in brain homeostasis (Lucas et al., 2010). The inflammatory response then shifts to the anti-inflammatory response, in which M2 microglia are responsible for clearing debris, angiogenesis, and extracellular matrix deposition and play a significant role in neuroprotection. When the pro-inflammatory response does not yield, the persistent presence and production of inflammatory cytokines and reactive oxygen species by M1 macrophages can lead to cell death and further tissue damages, including synapse changes. Furthermore, the inactivation of the M2 microglial response can also result in a prolonged state of inflammation in the CNS (Lucas et al., 2010). These conditions are associated with chronic inflammation in the CNS. Therefore, the failure in microglial phenotype switching is a potential contributor to neurodevelopmental disorders like ASD.

In a study conducted by Vargas et al. (2005) significant signs of microglial activation have been shown in different regions of post mortem brains in autistic patients, including the cortex, white matter, and

cerebellum, compared to brains of the members of the control group who had not any neurological disorders (Matta, et al., 2019). In this study, the signs of increased astrocyte activity in all areas, as mentioned earlier, marked by glial fibrillary acidic protein expression, have also been detected (Matta et al., 2019). These changes were most prominent in the cerebellum tissue (Matta et al., 2019).

According to another study, the density of microglia in the dorsolateral prefrontal sections of the brain cortex of autistic patients was significantly increased (Morgan et al., 2010). Another consistent study investigating the brain autopsies of individuals with autism found a marked increased microglial density in the fronto-insular (FI) and the visual cortex (VC) regions compared to neuro-normal individuals. Given that these two brain regions are entirely separate from each other, the authors of this study have suggested a generalized increase in microglial activity in the whole brain cortex of ASD patients (Tetreault et al., 2012). The difference in the autistic brain is not only confined to the quantity of microglia, but also the morphological parameters in microglia vary. For instance, despite a similar number of microglia in the brain amygdala in patients with ASD, the resident microglia in the amygdala were morphologically different compared to neuro-normal individuals (Morgan et al., 2014). Another study also demonstrated a significant elevation in the primed type of microglia and a remarkable decrease in the ramified type of microglia in the postmortem brains of patients with ASD compared to normal developing individuals despite the similar total density of the microglia (Lee et al., 2017).

One of the possible triggers of microglial activation in patients with ASD is the increased secretion of extracellular vesicles (EV) in these patients (Tsilioni and Theoharides, 2018). EVs, secreted from different cells, alter the function of target cells. In patients with ASD, they trigger microglial cells to secrete a higher amount of pro-inflammatory cytokines like IL1-B, a process that can ultimately lead to ASD development (Tsilioni and Theoharides, 2018).

The microbiome and gut-brain axis

The symptoms of gastrointestinal (GI) disturbances are relatively common among patients with ASD (Yang et al., 2018). One of the leading causes of these symptoms is intestinal microbiota imbalance, a condition called dysbiosis (Nitschke et al., 2020). The intestinal microbiota plays a vital role in the function and development of CNS, neuroendocrine, and neuroimmune systems (Nitschke et al., 2020). The gut-brain axis (GBA) is a bidirectional pathway between the CNS, autonomic

nervous system (ANS), the hypothalamic-pituitary-adrenal axis (HPA), and the enteric nervous system, enabling direct and indirect communication among them (Ferguson and Solo-Gabriele, 2016; Sivamaruthi et al., 2020). The sympathetic and parasympathetic nerves mediate efferent signals from the CNS to the intestine, regulating microbiome composition and transferring afferent signals from the intestine to the CNS (Sivamaruthi et al., 2020).

Besides, the microbiome transfers different messages *via* either secreting various neuro-active molecules such as acetylcholine or *via* activating the vagus nerve directly (Petra et al., 2015). Following environmental stresses and pro-inflammatory cytokine release caused by the change in the intestinal flora's composition, the HPA, responsible for emotional responses and memory, is activated, which ultimately results in cortisol secretion. Cortisol is capable of altering the brain, the intestinal endothelium, and the gut microbiota composition. Hence, the intestinal effector cells such as endothelium and immune cells are under CNS control by the hormonal and neuronal pathways. Besides, any dysregulation in the intestinal effector cells may lead to microbial composition and dysbiosis changes. This process can be followed by CNS development abnormalities and dysfunctions, including behavioral and neuropsychiatric diseases (Sivamaruthi et al., 2020). Moreover, a study conducted by Eltokhi et al. (2020), demonstrated that the microbiome has a critical role in the synaptic pruning process, specifically in the prenatal and early postnatal periods of life.

While discrepancies exist upon which bacteria are most responsible for dysbiosis, recently, the bacteria "Clostridium difficile" has been found to be significantly increased in the stool exam of individuals with ASD compared with normal controls (Saurman et al., 2020). Another study also revealed that the investigated autistic children with GI abnormalities had significantly higher levels of the intestinal bacteria "Clostridium perfringens" than neuro-normal children without GI symptoms (Finegold et al., 2017). However, no significant microbiota differences were found between autistic children and normal controls (Gondalia et al., 2012). These inconsistencies may be justified by the small sample sizes surveyed in these studies. The elucidation of the role of intestinal microbiome in patients with ASD can lead to better future interventions for relieving behavioral and cognitive autistic symptoms, including fecal microbiota transplantation, change in the diet, and utilization of probiotics and prebiotics (Eltokhi et al., 2020). For instance, dietary polyphenols were found to have promising effects on the manifestations of ASD by impacting the GBA (Serra et al., 2020).

Inflammatory and anti-inflammatory cytokines in individuals with ASD

Altered levels of cytokines are known as an essential component of immune dysregulation in patients with ASD. In a case-control study, the expression of the genes of IL-6, an inflammatory protein, and heat shock protein 70i (HSP70i), a stress-related protein, were significantly higher in children with ASD. Furthermore, the plasma levels of Prx2 and Prx5 (peroxiredoxins), antioxidant enzymes responsible for protecting the brain against oxidative stress, were significantly higher in children with ASD compared to typically developing children; signifying neuroinflammation as a part of autism pathogenesis (Abruzzo et al., 2019). Moreover, it has been reported that the granulocyte-macrophage colony-stimulating factor (GM-CSF), interferon- γ (IFN- γ), IL-6, IL-9, IL-22, T-bet, and phospho-signal transducer and activator of transcription-3 (pSTAT3) producing CD45 cells have been remarkably increased in the children with ASD compared with control subgroup (Ahmad et al., 2020).

It has also been shown that patients with ASD have decreased anti-inflammatory response of T helper-2 and the increased pro-inflammatory response of T helper-1 cells (Bjorklund et al., 2016).

In contrast to the mentioned studies, which demonstrated higher levels of inflammatory cytokines in patients with ASD, in a case-control study conducted by Gomez-Fernandez et al. (2018) no significant differences were observed in the levels of cytokines between the children with ASD and normal developing children, except for the nerve growth factor (NGF) which was higher in the autistic children. However, this study has shown lower plasma levels of neural cell adhesion molecule (NCAM) and higher NGF levels in ASD patients without developmental regression compared with the subgroup with developmental regression. This highlights the difference in pathological pathways among various subgroups of ASD (Gomez-Fernandez et al., 2018).

Moreover, in a meta-analysis, higher plasma levels of pro-inflammatory cytokines, including IFN- γ , IL-1 β , IL-6, and TNF- α were reported in patients with ASD compared to typically developing individuals (Saghazadeh et al., 2019b). Furthermore, significantly higher TNF- α and S100B (a calcium-binding protein) in autistic patients have been demonstrated in another study. Besides, the S100B peripheral level was higher in patients with severe ASD compared to those with mild to moderate forms. However, other pro-inflammatory cytokines were similar between the subjects with ASD and the ordinary individuals (Guloksuz et al., 2017).

One study addressed cytokine profiles in the siblings of ASD patients, including autistic children (N=80), unaffected siblings of the autistic children (51 persons), and unrelated normal controls (N=86). An increased plasma level of IL-6 was found in both autistic children and their siblings compared to the unrelated healthy control group. On the other hand, TNF- α and IL-8 were exclusively elevated in autistic children. However, the IL-9 and IL-10 levels did not show any differences between the mentioned three groups (Alzghoul et al., 2019).

The results of a meta-analysis showed that ASD patients have higher serum levels of IL-8 (Masi et al., 2015). Furthermore, IL-8 was the only cytokine that showed higher levels after adjusting parental cytokines in children with ASD, making IL-8 a promising biomarker for diagnosing ASD and contributing to the pathogenesis (Shen et al., 2020). In contrast, another study reported lower serum levels of IL-8, but higher brain levels were reported in patients with ASD (Businaro et al., 2016).

Besides, increased activity of superoxide dismutase (SOD) in the neutrophils and monocytes was found in patients with ASD compared to normal developing individuals. In comparison, the activities of glutathione peroxidase and glutathione reductase were reduced or unaltered in this population. This highlights the possible role of the dysregulated enzymatic antioxidant network in the pathogenesis of autism (Nadeem et al., 2019). On the other hand, a moderate reduction in the serum IL-10 levels, a little decreased serum IL-1 receptor antagonist, and a slightly increased plasma IL-5 level were found in patients with ASD (Saghazadeh et al., 2019a).

Although ASD has a strong genetic component, exposure to some environmental factors during pregnancy has been associated with offspring's autism-like behaviors. Valproic acid (VPA) is an anti-epileptic drug widely used in pregnancy (Zhao et al., 2019). The prenatal exposure to VPA in rodents has been associated with behavioral deficits consistent with ASD. This association was also approved in non-human primates in 2019 (Zhao et al., 2019). The VPA-induced animal model of autism was developed in 1996, and since then, this model has been extensively used to investigate different molecular pathways in ASD (Rodier et al., 1996). In an animal study, rats were intraperitoneally (IP) injected with VPA on day 12.5 of gestation, and then their inflammatory profile was investigated at 11 and 13 weeks of age. The VPA-induced rats showed significantly increased expression of pro-inflammatory cytokines – including IL-1B and TNF- α – and reduced expression of social behavior-related genes like BDNF and neuroligin-3 in the hippocampus compared

to control rats. Besides, the hippocampal expression of gamma-aminobutyric acid (GABA) and glutamic acid decarboxylase (GAD) was decreased (Win-Shwe et al., 2018). On the other hand, maternal immune activation in pregnancy-induced by polyinosinic-polycytidylic acid (poly (I:C)) is associated with autism in offspring (Lammert and Lukens, 2019). Poly (I:C) exposure in pregnancy has been associated with elevated immune response in several animal studies (Haddad, Patel, et al., Schmid, 2020). However, in a recent study on rats, the IP injection of poly (I:C) on gestational day 15 resulted in increased IL-6 level 6 h and 24 h post-injection but not in postnatal days (Murray et al., 2019). Lipopolysaccharide (LPS) administration in mice has also been connected to autism-like behaviors, and CNS changes consistent with autism (Custódio et al., 2018). In a study, Swiss mice were neonatally challenged with LPS in postnatal days (PN) 5-7 and analyzed in PN35 and PN70. The mice showed increased levels of IL-4 in the prefrontal cortex (PFC), hippocampus (HC), and hypothalamus (HT) and decreased levels of IL-6 in PFC, HC, and HT. Moreover, the BDNF levels were increased in both PN70 male and female mice. LPS-induced male mice showed increased myeloperoxidase (MPO) activity in both PN35 and PN70. At the same time elevated levels of IFN γ and nitrite and decreased parvalbumin were observed in PN70 male mice (Custódio et al., 2018). These observations have suggested a sex and age-specific alteration in cytokine profile resembling ASD following LPS-administration (Custódio et al., 2018).

Taken together, the available data on the profile of pro-inflammatory cytokines mainly indicated increased serum levels of IFN- γ , IL-1 β , IL-6, and TNF- α in patients with ASD. However, further investigations on the pro-inflammatory and anti-inflammatory cytokines in autistic patients with adequate sample sizes are recommended.

The family history of immune dysregulation in children with ASD

Concerning the possible involvement of autoimmunity in the pathogenesis of ASD and the inheritable nature of most autoimmune disorders, several studies have investigated autoimmune diseases in the family members of autistic children. A multi-site case-control study investigated the presence of asthma, any autoimmune diseases, and allergies in the family members of autistic children. The mothers with a past medical history of asthma were significantly more likely to have a child with autism than the mothers ascertained from the general population ($OR=1.26$, $P=0.05$). Besides, the

prevalence of other autoimmune disorders and allergies was higher in autistic children's mothers, although the differences were not significant. Moreover, this study reported that maternal immune conditions in pregnancy are significantly followed by developmental disorders in the child ($OR=1.37$, $P=0.03$) but not autism alone ($OR=1.29$, $P=0.15$). However, no significant association was found between paternal autoimmune diseases and the development of autism in children (Croen et al., 2019).

In a cohort study, a significant association was found between infantile autism and family history of type 1 diabetes mellitus (T1DM). Besides, a significant association was reported between maternal history of rheumatoid arthritis (RA) and celiac disease and the presence of ASD in the children (Atladóttir et al., 2009). Moreover, this study showed that the association between the family history of RA and ASD in the child was limited to maternal history, compared with an older study that showed a higher prevalence of ASD in children with an overall family history of RA (Comi et al., 1999). These conflicting results may emphasize an altered antibody exposure in the prenatal period in children with ASD (Atladóttir et al., 2009).

To shed light on the specific factor leading to ASD in children and immune dysregulation in their family members, Mostafa and Shehab (2010) designed a study and proposed that the complement 4B null allele has a significant role in ASD. They demonstrated that C4B null allele was significantly more frequent in children with ASD compared to the control group ($P<0.001$) and also had a significant association with both ASD ($OR=6.26$, 95% $CI=2.5-14.1$) in the children and their family history of autoimmunity ($OR=21$, 95% $CI=2.5-14.1$). Besides, the family history of autoimmune abnormalities in children with ASD was significantly more frequent than the standard developing control group ($P<0.001$) (Mostafa and Shehab, 2010).

Furthermore, regressive autism, which is defined as the loss of speech and social skills several months after birth, was associated with a family history of autoimmune diseases ($OR=1.89$) (Molloy et al., 2006). The authors also found that autoimmune thyroid disease was the only autoimmune disorder in family members of autistic children to have a significant association with the regressive type of ASD in the child ($OR=2.09$, $P=0.003$). It was also shown that children with more than one family member affected with autoimmune diseases were more likely to be diagnosed with regressive ASD rather than non-regressive type ($OR=1.89$, $P=0.009$) (Molloy et al., 2006). Besides, maternal hypothyroidism diagnosed and treated for the first time after the child's birth was significantly associated with the presence of autism in offsprings. The abnormal

level of thyroid hormones in mothers with untreated hypothyroid during pregnancy may result in neuro-developmental disorders in the child, including ASD (Brown et al., 2015).

Besides, according to an animal study, extreme maternal hypothyroxinemia was associated with a four times greater probability of having a child with autism (adjusted $OR=3.89$, $P<0.001$) (Román, 2013). However, another study reported that maternal autoimmune diseases were significantly associated with developmental disorders in their offsprings ($OR=1.46$) but not exclusively with autism (Lyall et al., 2014). Moreover, another study reported a higher frequency of maternal immunological abnormalities in 4 years around pregnancy. However, it did not vary significantly between the autistic children and the control group (10.3% vs. 8.2%, $P=0.15$), except for maternal psoriasis, which was significantly associated with developing ASD ($p=0.003$, $OR=2.7$) (Croen et al., 2005). Furthermore, according to Mouridsen et al. (2007) infantile autism (IA) was significantly associated with maternal ulcerative colitis ($P=0.05$) and paternal T1DM ($P=0.02$). However, they found no differences in the prevalence of autoimmune diseases between mothers of autistic children and mothers recruited from the general population ($P=0.71$). The prevalence of autoimmune diseases in fathers in the case and control group was 8.6% versus 4.6%, respectively ($P=0.14$). Besides, Sweeten et al. (2003) showed that the frequency of autoimmune abnormalities was significantly higher in the family members of children with pervasive developmental disorders, a term used to describe developmental disorders, including autism in the DSM-IV definition. Despite some discrepancies, some autoimmune disorders in the family such as T1DM, hypothyroidism, RA, psoriasis, and ulcerative colitis are associated with the development of ASD in children. However, there is still a need for more research in this field.

Known concomitant autoimmune diseases in patients with ASD

Up to the present, the co-occurrence of some autoimmune disorders, including T1DM, hypothyroidism, celiac disease, and inflammatory bowel disease with ASD, has been investigated. The prevalence of ASD among children with T1DM who attended a diabetic clinic in Toronto was greater than the general population (Freeman et al., 2005). In line with this study, it has been clarified that the prevalence of ASD in T1DM patients in Colorado and Ontario were 1.16% and 0.9%, respectively, both higher than the ASD prevalence

in the normal population living in Colorado (0.7%) (Stanek et al., 2019). The same authors found a lower HbA1c level ($P<0.0001$) and insulin pump ($P<0.0001$) in ASD+T1DM patients compared to T1DM patients. However, several studies examined a higher prevalence of T1DM in ASD patients, including a survey among children with pre-diagnosed T1DM ($N=10032$), which indicated no difference in the prevalence of ASD between these children and normal children. They have also reported lower HbA1c concentrations and lower pump use in children with ASD, which may be due to the more regimented routines in this population (Bethin et al., 2019). Moreover, no higher prevalence of ASD was found in children with T1DM ($N=5178$) in Finland (Harjutsalo and Tuomilehto, 2006).

Besides, no connection has been found between the overall ASD prevalence and the increased blood levels of TSH as a sign of hypothyroidism in the early life period. However, it has been suggested that the high TSH level may have an association with the regressive sub-phenotype of ASD (Ames et al., 2020).

Although case reports have shown celiac disease and ASD's co-occurrence, other studies did not support this finding (Ludvigsson et al., 2013; Bavykina et al., 2018). The predominant gluten sensitivity was reported in 41.9% of ASD children; however, no beneficiary role of a gluten-free diet was found in children with ASD (Buie, 2013). Other studies which reported no relationship between ASD and celiac disease were conducted on small sample sizes (Juneja et al., 2018; Batista et al., 2012; Pavone et al., 1997). In a registry in Sweden, patients with celiac disease (27000) showed no association between prior ASD and celiac disease. An apparent increased risk of ASD was also reported in patients with normal mucosa but with positive celiac auto-antibodies ($OR=4.57$; 95% $CI=1.58-13.22$) (Ludvigsson et al., 2013).

As pointed out in the previous sections, ASD pathogenesis may be associated with microbiome dysregulation and the presence of inflammation in the bowels. In line with this, a retrospective case-cohort study indicated that children with ASD had a higher prevalence of Crohn's disease and ulcerative colitis than the control group (Lee et al., 2018).

Anti-inflammatory treatment in relieving the symptoms of ASD

As mentioned earlier, there is recently growing literature pointing out the critical role of inflammation in the pathogenesis of autism. Accordingly, anti-inflammation therapies have much been investigated for relieving autism manifestations. Some anti-in-

flammatory interventions, which have been studied in patients with autism, are sulforaphane, osthole, fexofenadine, resveratrol, palmitoylethanolamide (PEA), pioglitazone, acetylcysteine, propentofylline, L-carnosine, yokukansan, spironolactone, celecoxib, flavonoid luteolin, corticosteroids, minocycline, and stem cell therapy.

Sulforaphane is found chiefly in cruciferous vegetables like broccoli, sulforaphane is known for its anti-inflammatory and antioxidant effects (Durham et al., 2014). In a placebo-controlled randomized trial among men aged 13 to 27 years with ASD (N=44), sulforaphane significantly improved autistic behaviors compared with the placebo group. In other words, it led to a substantial decrease in the scores of the Aberrant Behavior Checklist ($p < 0.001$), social responsiveness scale ($p < 0.017$), and clinical global impression improvement scale ($P = 0.007-0.015$) (Singh et al., 2014).

Osthole is used in Chinese traditional medicine and has been reported to have anti-inflammatory effects (Zhang et al., 2015). Similarly, fexofenadine is a known anti-histamine drug with anti-inflammatory effects (Kordulewska et al., 2019). According to Kordulewska et al. (2019) the cyclooxygenase 2 (COX-2) pathways may have a role in the pathogenesis of autism, and osthole or fexofenadine application in children with autism can decline the COX-2 inflammatory effects. It may lead to a reduction in autism development.

Resveratrol (RSV) is a polyphenolic substance, which can be obtained from some plants and fruits and recently has been extensively the subject of research for its numerous beneficial effects, including antioxidant and anti-inflammatory effects (Gambini et al., 2015). Resveratrol acts by affecting apoptosis of activated T cells and suppressing TNF- α , IL-17, and the other pro-inflammatory cytokines (Diaz-Gerevini et al., 2016). An animal study showed resveratrol has promising effects on restoring all the core and associated manifestations of autism by inhibiting oxidative-nitrosative stress in the rats (Bhandari and Kuhad, 2017). Moreover, resveratrol improved defective mitochondrial fatty acid oxidation in patients with autism (Barone et al., 2019). However, no randomized control trial has been conducted up to date on resveratrol effects on patients with ASD.

Palmitoylethanolamide, an endocannabinoid molecule, has potential anti-inflammatory effects (Khalaj et al., 2018). It significantly improved autism-related irritability and hyperactivity symptoms when adjunct to risperidone therapy (Khalaj et al., 2018). Consistently, it improved autistic-like behaviors by affecting intestinal microbial composition in mice (Cristiano et al., 2018).

Pioglitazone is a member of thiazolidinedione drugs, widely used as an anti-diabetic agent with anti-inflammatory effects. Daily therapy with 30-60 mg of pioglitazone in children with autism (N=30) has led to apparent clinical improvements in irritability, lethargy, stereotypy, and hyperactivity subscales of autism (Boris et al., 2007). Moreover, in an animal study, daily pioglitazone use in rats, which were induced by lipopolysaccharide in the prenatal period, improved their autistic-like behaviors and abolished their IL-6 levels (Kirsten et al., 2018).

Acetylcysteine is known for its antioxidant effects and has recently-demonstrated anti-inflammatory properties (Uraz et al., 2013). Acetylcysteine could reduce the aggressive and unpredictable behaviors in a 17-year-old boy who has autism, who did not respond to the other medications (Stutzman and Dopheide, 2015).

L-carnosine belongs to the family of hybrid peptides with reported anti-inflammatory and anti-oxidation effects (Tsai et al., 2010). In a randomized, double-blind placebo-controlled study, children with autism (N=70) were enrolled and randomly assigned to 10-week therapy with placebo plus risperidone or L-carnosine plus risperidone regimen. L-carnosine subgroup had better scores in the hyperactivity-non-compliance subscale of the Aberrant Behavior Checklist-Community rating scale (Hajizadeh-Zaker et al., 2018). However, this study reported no significant differences in the irritability subscale between groups (Hajizadeh-Zaker et al., 2018).

Spironolactone is a potassium-sparing diuretic, which has anti-androgenic effects (Marchezan et al., 2018). It has also shown its potential to be used as an anti-inflammatory drug by inhibiting the production of inflammatory cytokines such as TNF- α (Bendtsen et al., 2003). A case-report study found that spironolactone had promising effects on autistic behaviors and ABC scores of a 12-year-old autistic boy (Bradstreet et al., 2007).

Flavonoid luteolin can be found in plenty of plants. It has been demonstrated that it can inhibit the secretion of pro-inflammatory cytokines from mast cells in humans (Kempuraj et al., 2005). In an uncontrolled case series, the use of luteolin for at least four months by children with autism (N=37) improved GI and allergy symptoms in 75%, attention and eye contact in 50%, and social interaction in 25% of them (Theoharides et al., 2012). In line with this study, an open-label trial reported the beneficiary roles of combined flavonoid luteolin and quercetin in relieving autism symptoms (Taliou et al., 2013). Moreover, a significant improvement was reported in autistic behaviors in one mouse model following the combined administration of luteolin and palmitoylethanolamide (Bertolino et al., 2017).

Table 1. Anti-inflammatory therapies among subjects with ASD.

Therapy	Methodology	Measure	Sample size	Dosage	Period	Outcomes	References
Sulforaphane	Placebo-controlled, double-blind, randomized trial	ABC ¹ -SRS ² -CGI-I ³	29	50–150 µmol	18 weeks	Decline in scores: 34% for ABC, 17% for SRS n CGI-I: SI, VC and AB improvements	(Singh et al., 2014)
(Palmitoylethanolamide (PEA) + risperidone) versus (risperidone + placebo)	Randomized, parallel-group, double-blind placebo-controlled trial	ABC-C ⁴	70	600 mg PEA twice daily	10 weeks	Significant improvements in ABC-irritability and hyperactivity/noncompliance symptoms (p=0.001), great effect on inappropriate speech (p=0.051)	(Khalaj et al., 2018)
L-carnosine add on to risperidone	Randomized, double-blind, placebo-controlled trial	ABC-C - SORS	70	800 mg/day l-carnosine in 2 divided doses	10 weeks	Significant improvement in hyperactivity/noncompliance subscale (p=0.044)	(Hajizadeh-Zaker et al., 2018)
Minocycline as an adjunctive to risperidone	Randomized controlled trial	ABC-C	46	50 mg twice per day	10 weeks	Significant improvement in irritability, hyperactivity/noncompliance	(Ghaleiha et al., 2016)
Celecoxib as an adjunctive to risperidone	Randomized double-blind placebo-controlled	ABC-C	40	300 mg/day	100 weeks	Significant improvement in Irritability, Lethargy/Social Withdrawal and Stereotypic Behavior	(Asadabadi et al., 2013)
Flavonoid luteolin + flavonoid quercetin	Prospective, open-label trial	VABS ⁵ -ABC - CGI-I - Autism Treatment Evaluation Checklist	50	1 capsule (100 mg luteolin, 70 mg quercetin) per 10 kg weight per day	26 weeks	Reduction (26.6%-34.8%) in Aberrant Behavior Checklist subscale scores- significant improvement in adaptive functioning measured by VABS- Transient increased irritability (1-8w) in 27 subjects	(Taliou et al., 2013)
Pioglitazone	Small cohort	ABC	25	30 mg (age 3-5) or 60 mg (age 6-17) daily	3-4 months	Significant improvement in irritability, lethargy, stereotypy, and hyperactivity	(Boris et al., 2007)
Flavonoid luteolin + flavonoid quercetin	Uncontrolled open case series	Observations of the responses by the parents of children with ASD	37	2 capsules/ 20 kg weight or at least 400 mg total flavonoid	at least 4 months	75% improvement in GI and allergy symptoms, 50% in eye contact and attention, 25% in social interaction, 10% in a resumption of speech	(Theoharides et al., 2012)
Acetylcysteine	Case report	Occupational and recreation therapists observations	1	600 mg twice daily	6 weeks	Reduction in the patient's aggressive behavior, tantrums, and irritability	(Stutzman and Dopheide, 2015)

Therapy	Methodology	Measure	Sample size	Dosage	Period	Outcomes	References
Spirolactone	Case report	ABC	1	2 mg/kg daily	4 weeks	79% improvement in irritability, 83% decrease in lethargy, a 60% reduction of stereotypy, 72% reduction of hyperactivity, and a 67% decrease in inappropriate speech	(Bendtsen et al., 2003)
Corticosteroids	Case report	Observation and WISC-R ⁶	1	Begin with 2 mg/kg prednisolone, titrated monthly by 0.5 mg/kg from weeks 4 through 12	28 weeks	Amelioration of language abilities and behavior and increase in IQ, increase in spontaneous speech, greater responsiveness to verbal communications, and improved social relatedness	(Stefanatos et al., 1995)
Resveratrol	Animal study	-	-	5, 10 and 15 mg/kg	4 weeks	Restore all the neurological, sensory, behavioral, biochemical and molecular deficits in autistic rats	(Bhandari and Kuhad, 2017)
Palmitoylethanolamide (PEA)	Animal study	Serum parameters, mitochondrial parameters, Self-Grooming test, Marble burying assay	-	10 or 30 mg/kg	10 days	Improvement in autistic-like behaviors in mice, mitochondrial dysfunction, reduction in the overall inflammatory state and expression of pro-inflammatory cytokines	(Cristiano et al., 2018)
Pioglitazone	Animal study	Plasma evaluation, play behavior evaluation, vocalization evaluation	6-8 rats	0.25 mg/kg/day	10 days	Reduction in IL-6 level, improvement in social interaction and the number of vocalizations	(Kirsten et al., 2018)

1 Aberrant Behavior Checklist.

2 Social Responsiveness Scale.

3 Clinical Global Impression – Improvement scale.

4 Aberrant behavior checklist – community.

5 Vineland Adaptive Behavior Scales.

6 Wechsler Intelligence Scale for Children-Revised.

Corticosteroids are a well-known group of anti-inflammatory drugs, widely used to treat plenty of disorders. A case report showed improved language abilities and autistic behaviors in a 6-year-old child with a pervasive developmental disorder (by the definition of DSM-IV) (Stefanatos et al., 1995). Furthermore, two childhood disintegrative disorder cases showed a significant improvement in behavior, motor regression,

and language following corticosteroid therapy (Mordekar et al., 2009).

Minocycline is an antibiotic whose neuroprotective and anti-inflammatory effects have been explored recently (Elewa et al., 2006). In a randomized control trial conducted by Ghaleliha et al. (2016) forty-six children with ASD were recruited and entered in 10-week risperidone plus minocycline or risperidone plus placebo

treatment groups. The children receiving minocycline as an adjunctive therapy had better scores in ABC-C irritability ($P=0.02$) and hyperactivity/noncompliance ($P=0.002$) sub-scales. However, there were no significant differences in terms of social withdrawal, stereotypical behaviors, and inappropriate speech sub-scales of ASD between minocycline and placebo groups (Ghaleiha et al., 2016).

Celecoxib is a non-steroidal anti-inflammatory drug (Shin, 2018). Celecoxib was investigated in a 10-week randomized control trial among children with ASD ($N=40$). They entered either risperidone plus celecoxib or risperidone plus placebo treatment groups. The results indicated that children who received a celecoxib-added regimen had significant improvement in the subscales of irritability ($P<0.001$), lethargy/social withdrawal ($P<0.001$), and stereotypic behaviors ($P<0.001$) compared with the placebo group. Moreover, the rate of adverse effects was similar between the two groups (Asadabadi et al., 2013).

Stem cell therapy alleviates the symptoms of autism by an unknown molecular pathway. One explanation for its effects is the paracrine activity by stem cells, by which plenty of anti-inflammatory cytokines, including exosomes are secreted (Alessio et al., 2020).

CONCLUSION

Taken together, autoimmunity may be an essential nominee for the pathogenesis of ASD. The involvement of microglia and astrocytes in the process of CNS inflammation, the role of the gut-brain axis in the CNS development, the presence of higher pro-inflammatory cytokines in patients with ASD, the higher prevalence of immune dysregulation in the family members of affected children, and the successful application of some anti-inflammatory therapies in patients with ASD are some pieces of evidence to count ASD as a neuroimmune disorder. However, inconsistencies exist, and there are still many unanswered questions. Thus, further studies with robust methodologies should be a priority.

REFERENCES

- Abruzzo PM, Matté A, Bolotta A, Federti E, Ghezzi A, Guarnieri T, Marini M, Posar A, Siciliano A, De Franceschi L, Visconti P (2019) Plasma peroxiredoxin changes and inflammatory cytokines support the involvement of neuroinflammation and oxidative stress in autism spectrum disorder. *J Translational Med* 17: 332.
- Ahmad SF, Ansari MA, Nadeem A, Bakheet SA, Al-Ayadhi LY, Alasmari AF, Alanazi MM, Al-Mazroua HA, Attia SM (2020) Involvement of CD45 cells in the development of autism spectrum disorder through dysregulation of granulocyte-macrophage colony-stimulating factor, key inflammatory cytokines, and transcription factors. *Int Immunopharmacol* 83: 106466.
- Alessio N, Brigida AL, Peluso G, Antonucci N, Galderisi U, Siniscalco D (2020) Stem cell-derived exosomes in autism spectrum disorder. *Int J Environmental Res Public Health* 17: 944.
- Allen NJ (2014) Astrocyte regulation of synaptic behavior. *Annu Rev Cell Dev Biol* 30: 439–463.
- Alzghoul L, Abdelhamid SS, Yanis AH, Qwaider YZ, Aldahabi M, Albodour SA (2019) Association between levels of inflammatory markers in autistic children compared to their unaffected siblings and unrelated healthy controls. *Turk J Med Sci* 49: 1047–53.
- Ames JL, Windham GC, Lyall K, Pearl M, Kharrazi M, Yoshida CK, Van de Water J, Ashwood P, Croen LA (2020) Neonatal thyroid stimulating hormone and subsequent diagnosis of autism spectrum disorders and intellectual disability. *Autism Res* 13: 444–455.
- Araque A, Parpura V, Sanzgiri RP, Haydon PG (1999) Tripartite synapses: glia, the unacknowledged partner. *Trends Neurosci* 22: 208–215.
- Asadabadi M, Mohammadi MR, Ghanizadeh A, Modabbernia A, Ashrafi M, Hassanzadeh E, Forghani S, Akhondzadeh S (2013) Celecoxib as adjunctive treatment to risperidone in children with autistic disorder: a randomized, double-blind, placebo-controlled trial. *Psychopharmacology* 225: 51–59.
- American Psychiatric Association (2013) Diagnostic and statistical manual of mental disorders (DSM-5®) (American Psychiatric Pub).
- Atladóttir HO, Pedersen MG, Thorsen P, Mortensen PB, Deleuran B, Eaton WW, Parner ET (2009) Association of family history of autoimmune diseases and autism spectrum disorders. *Pediatrics* 124: 687–694.
- Baio J, Wiggins L, Christensen DL, Maenner MJ, Daniels J, Warren Z, Kurzius-Spencer M, Zahorodny W, Robinson Rosenberg C, White T, Durkin MS, Imm P, Nikolaou L, Yeargin-Allsopp M, Lee LC, Harrington R, Lopez M, Fitzgerald RT, Hewitt A, Pettygrove S, Constantino JN, Vehorn A, Shenouda J, Hall-Lande J, Van Naarden Braun K, Dowling NF (2014) Prevalence of autism spectrum disorder among children aged 8 years—autism and developmental disabilities monitoring network, 11 sites, United States, 2010. *MMWR Surveill Summ* 67: 1–23.
- Barone R, Rizzo R, Tabbi G, Malaguarnera M, Frye RE, Bastin J (2019) Nuclear peroxisome proliferator-activated receptors (PPARs) as therapeutic targets of resveratrol for autism spectrum disorder. *Int J Mol Sci* 20: 1878.
- Batista IC, Gandolfi L, Nobrega YK, Almeida RC, Almeida LM, Campos Junior D, Pratesi R (2012) Autism spectrum disorder and celiac disease: no evidence for a link. *Arq Neuropsiquiatr* 70: 28–33.
- Bavykina IA, Zvyagin AA, Petrova IV, Nastaushva TL (2018) Markers of gluten intolerance in children with autism spectrum disorders and Down syndrome (in Russian). *Zh Nevrol Psikhiatr Im S S Korsakova* 118: 64–68.
- Bendtsen K, Hansen PR, Rieneck K, Spironolactone/Arthritis Study Group (2003) Spironolactone inhibits the production of proinflammatory cytokines, including tumour necrosis factor- α and interferon- γ , and has potential in the treatment of arthritis. *Clin Exp Immunol* 134: 151–158.
- Bertolino B, Crupi R, Impellizzeri D, Bruschetta G, Cordaro M, Siracusa R, Esposito E, Cuzzocrea S (2017) Beneficial effects of co-ultramicrosized palmitoylethanolamide/luteolin in a mouse model of autism and in a case report of autism. *CNS Neurosci Ther* 23: 87–98.
- Bethin KE, Kanapka LG, Laffel LM, Majidi S, Chaytor NS, MacLeish S, Adams RA, Foster NC, T1D Exchange Clinic Network (2019) Autism spectrum disorder in children with type 1 diabetes. *Diabet Med* 36: 1282–1286.
- Bhandari R, Kuhad A (2017) Resveratrol suppresses neuroinflammation in the experimental paradigm of autism spectrum disorders. *Neurochem Int* 103: 8–23.
- Bilimoria PM, Stevens B (2015) Microglia function during brain development: new insights from animal models. *Brain Res* 1617: 7–17.
- Bitzer-Quintero OK, González-Burgos I (2012) Immune system in the brain: a modulatory role on dendritic spine morphophysiology? *Neural Plast* 2012: 348642.

- Bjorklund G, Meguid NA, El-Bana MA, Tinkov AA, Saad K, Dadar M, Hemimi M, Skalny AV, Hosnedlová B, Kizek R, Osredkar J, Urbina MA, Fabjan T, El-Houfey AA, Kažužna-Czaplińska J, Gałtarek P, Chirumbolo S (2020) Oxidative stress in autism spectrum disorder. *Mol Neurobiol* 57: 2314–2332.
- Bjorklund G, Saad K, Chirumbolo S, Kern JK, Geier DA, Geier MR, Urbina MA (2016) Immune dysfunction and neuroinflammation in autism spectrum disorder. *Acta Neurobiol Exp* 76: 257–268.
- Blanco-Suárez E, Caldwell AL, Allen NJ (2017) Role of astrocyte-synapse interactions in CNS disorders. *J Physiol* 595: 1903–1916.
- Boris M, Kaiser CC, Goldblatt A, Elice MW, Edelson SM, Adams JB, Feinstein DL (2007) Effect of pioglitazone treatment on behavioral symptoms in autistic children. *J Neuroinflammation* 4: 3.
- Bradstreet JJ, Smith S, Granpeesheh D, El-Dahr JM, Rossignol D (2007) Spironolactone might be a desirable immunologic and hormonal intervention in autism spectrum disorders. *Med Hypotheses* 68: 979–987.
- Brown AS, Surcel HM, Hinkka-Yli-Salomäki S, Cheslack-Postava K, Bao Y, Sourander A (2015) Maternal thyroid autoantibody and elevated risk of autism in a national birth cohort. *Prog Neuropsychopharmacol Biol Psychiatry* 57: 86–92.
- Buie T (2013) The relationship of autism and gluten. *Clin Therapeutics* 35: 578–583.
- Businaro R, Corsi M, Azzara G, Di Raimo T, Laviola G, Romano E, Ricci L, Maccarrone M, Aronica E, Fuso A (2016) Interleukin-18 modulation in autism spectrum disorders. *J Neuroinflammation* 13: 1–13.
- Cao F, Yin A, Wen G, Sheikh AM, Tauqeer Z, Malik M, et al. (2012) Alteration of astrocytes and Wnt/ β -catenin signaling in the frontal cortex of autistic subjects. *J Neuroinflamm* 9: 223.
- Cherry JD, Olschowka JA, O'Banion MK (2014) Neuroinflammation and M2 microglia: the good, the bad, and the inflamed. *J Neuroinflamm* 11: 98.
- Chever O, Djukic B, McCarthy KD, Amzica F (2010) Implication of Kir4.1 channel in excess potassium clearance: an in vivo study on anesthetized glial-conditional Kir4.1 knock-out mice. *J Neurosci* 30: 15769–15777.
- Chung WS, Allen NJ, Eroglu C (2015) Astrocytes control synapse formation, function, and elimination. *Cold Spring Harb Perspect Biol* 7: a020370.
- Chung EH, Chou J, Brown KA (2020) Neurodevelopmental outcomes of preterm infants: a recent literature review. *Transl Pediatrics* 9: S3.
- Comi AM, Zimmerman AW, Frye VH, Law PA, Peeden JN (1999) Familial clustering of autoimmune disorders and evaluation of medical risk factors in autism. *J Child Neurol* 14: 388–394.
- Cristiano C, Pirozzi C, Coretti L, Cavaliere G, Lama A, Russo R, et al. (2018) Palmitoylethanolamide counteracts autistic-like behaviours in BTBR T+ tf/J mice: Contribution of central and peripheral mechanisms. *Brain Behav Immun* 74: 166–175.
- Croen LA, Grether JK, Yoshida CK, Odouli R, Van de Water J (2005) Maternal autoimmune diseases, asthma and allergies, and childhood autism spectrum disorders: a case-control study. *Arch Pediatrics Adolesc Med* 159: 151–157.
- Croen LA, Qian Y, Ashwood P, Daniels JL, Fallin D, Schendel D, Schieve LA, Singer AB, Zerbo O (2019) Family history of immune conditions and autism spectrum and developmental disorders: Findings from the study to explore early development. *Autism Res* 12: 123–35.
- Custódio CS, Mello BS, Filho A, de Carvalho Lima CN, Cordeiro RC, Miyajima F, et al. (2018) Neonatal immune challenge with lipopolysaccharide triggers long-lasting sex- and age-related behavioral and immune/neurotrophic alterations in mice: relevance to autism spectrum disorders. *Mol Neurobiol* 55: 3775–3788.
- Diaz-Gerevini GT, Repossi G, Dain A, Tarres MC, Narasimha Das U, Eynard AR (2016) Beneficial action of resveratrol: how and why? *Nutrition* 32: 174–78.
- Dunn AJ (2001) Nervous and immune system interactions. e LS doi.org/10.1038/npng.els.0000195.
- Durham A, Jazrawi E, Rhodes JA, Williams C, Kilty I, Barnes P, Chung KF, Adcock I (2014) The anti-inflammatory effects of sulforaphane are not mediated by the Nrf2 pathway. *Eur Respirat J* 44: P3332.
- Edmiston E, Ashwood P, Van de Water J (2017) Autoimmunity, autoantibodies, and autism spectrum disorder. *Biol Psychiatry* 81: 383–90.
- Elewa HF, Hilali H, Hess DC, Machado LS, Fagan SC (2006) Minocycline for short-term neuroprotection. *Pharmacotherapy* 26: 515–521.
- Eltokhi A, Janmaat IE, Genedi M, Haarman BCM, Sommer IEC (2020) Dysregulation of synaptic pruning as a possible link between intestinal microbiota dysbiosis and neuropsychiatric disorders. *J Neurosci Res* 98: 1335–1369.
- Ferguson A, Solo-Gabriele H (2016) Children's exposure to environmental contaminants: an editorial reflection of articles in the IJERPH special issue entitled "Children's exposure to environmental contaminants." In: *Multidisciplinary Digital Publishing Institute*.
- Finegold SM, Summanen PH, Downes J, Corbett K, Komoriya T (2017) Detection of *Clostridium perfringens* toxin genes in the gut microbiota of autistic children. *Anaerobe* 45: 133–137.
- Fombonne E (2005) Epidemiological studies of pervasive developmental disorders. In: *Handbook of autism and pervasive developmental disorders: Diagnosis, development, neurobiology, and behavior* (Volkmar FR, Paul R, Klin A, Cohen D (Eds.). John Wiley & Sons Inc., p. 42–69.
- Freeman SJ, Roberts W, Daneman D (2005) Type 1 diabetes and autism: Is there a link? *Diabetes Care* 28: 925–26.
- Freeman MR (2010) Specification and morphogenesis of astrocytes. *Science* 330: 774–778.
- Gambini J, Inglés M, Olaso G, Lopez-Grueso R, Bonet-Costa V, Gimeno-Mallench L, Mas-Bargues C, Abdelaziz KM, Gomez-Cabrera MC, Vina J, Borras C (2015) Properties of resveratrol: in vitro and in vivo studies about metabolism, bioavailability, and biological effects in animal models and humans. *Oxid Med Cell Longev* 2015: 837042.
- Ghaleiha A, Alikhani R, Kazemi MR, Mohammadi MR, Mohammadinejad P, Zeinoddini A, Hamed M, Shahriari M, Keshavarzi Z, Akhondzadeh S (2016) Minocycline as adjunctive treatment to risperidone in children with autistic disorder: a randomized, double-blind placebo-controlled trial. *J Child Adolesc Psychopharmacol* 26: 784–791.
- Ghaleiha A, Alikhani R, Kazemi MR, Mohammadi M-R, Mohammadinejad P, Zeinoddini A, et al. (2016) Minocycline as adjunctive treatment to risperidone in children with autistic disorder: a randomized, double-blind placebo-controlled trial. *J Child Adolesc Psychopharmacology* 26: 784–791.
- Gomez Perdiguero E, Klapproth K, Schulz C, Busch K, Azzone E, Crozet L, et al. (2015) Tissue-resident macrophages originate from yolk-sac-derived erythro-myeloid progenitors. *Nature* 518: 547–551.
- Gomez-Fernandez A, de la Torre-Aguilar MJ, Gil-Campos M, Flores-Rojas K, Cruz-Rico MD, Martin-Borreguero P, Perez-Navero JL (2018) Children with autism spectrum disorder with regression exhibit a different profile in plasma cytokines and adhesion molecules compared to children without such regression. *Front Pediatrics* 6: 264.
- Gondalia SV, Palombo EA, Knowles SR, Cox SB, Meyer D, Austin DW (2012) Molecular characterisation of gastrointestinal microbiota of children with autism (with and without gastrointestinal dysfunction) and their neurotypical siblings. *Autism Res* 5: 419–427.
- Gordon S, Mantovan A (2011) Diversity and plasticity of mononuclear phagocytes. *Eur J Immunol* 41: 2470–2472.
- Gottfried C, Bambini-Junior V, Francis F, Riesgo R, Savino W (2015) The impact of neuroimmune alterations in autism spectrum disorder. *Front Psychiatry* 6: 121.
- Guillamón-Vivancos T, Gómez-Pinedo U, Matías-Guiu J (2015) Astrocytes in neurodegenerative diseases (I): function and molecular description. *Neurologia* 30: 119–129.
- Guloksuz SA, Abali O, Cetin EA, Bilgic Gazioglu S, Deniz G, Yildirim A, Kawikova I, Guloksuz S, Leckman JF (2017) Elevated plasma concentrations of S100 calcium-binding protein B and tumor necrosis factor- α in children with autism spectrum disorders. *Braz J Psychiatry* 39: 195–200.
- Habibi L, Ebtekar M, Jameie SB (2009) Immune and nervous systems share molecular and functional similarities: memory storage mechanism. *Scand J Immunol* 69: 291–301.

- Haddad FL, Patel SV, Schmid S (2020) Maternal immune activation by poly I:C as a preclinical model for neurodevelopmental disorders: A focus on autism and schizophrenia. *Neurosci Biobehav Rev* 113: 546–567.
- Hajizadeh-Zaker R, Ghajar A, Mesgarpour B, Afarideh M, Mohammadi MR, Akhondzadeh S (2018) l-carnosine as an adjunctive therapy to risperidone in children with autistic disorder: A randomized, double-blind, placebo-controlled trial. *J Child Adolesc Psychopharmacol* 28: 74–81.
- Hansen SN, Schendel DE, Parner ET (2015) Explaining the increase in the prevalence of autism spectrum disorders: the proportion attributable to changes in reporting practices. *JAMA Pediatrics* 169: 56–62.
- Harjutsalo V, Tuomilehto J (2006) Type 1 Diabetes and Autism: Is there a link?: Response to Freeman et al. *Diabetes Care* 29: 484–485.
- Juneja M, Venkatakishnan A, Kapoor S, Jain R (2018) Autism spectrum disorders and celiac disease: Is there an association? *Indian Pediatr* 55: 912–914.
- Kaufmann WE, Kidd SA, Andrews HF, Budimirovic DB, Esler A, Haas-Givler B, Stackhouse T, Riley C, Peacock G, Sherman SL (2017) Autism spectrum disorder in fragile X syndrome: co-occurring conditions and current treatment. *Pediatrics* 139: S194–S206.
- Kempuraj D, Madhappan B, Christodoulou S, Boucher W, Cao J, Papadopoulou N, Cetrulo CL, Theoharides TC (2005) Flavonols inhibit proinflammatory mediator release, intracellular calcium ion levels and protein kinase C theta phosphorylation in human mast cells. *Br J Pharmacol* 145: 934–944.
- Kettenmann H, Hanisch UK, Noda M, Verkhratsky AI (2011) Physiology of microglia. *Physiological Rev* 91: 461–553.
- Khalaj M, Saghadzadeh A, Shirazi E, Shalabafan MR, Alavi K, Shooshtari MH, et al. (2018) Palmitoylethanolamide as adjunctive therapy for autism: Efficacy and safety results from a randomized controlled trial. *J Psychiatry Res* 103: 104–111.
- Kim HJ, Cho MH, Shim WH, Kim JK, Jeon EY, Kim DH, Yoon SY (2017) Deficient autophagy in microglia impairs synaptic pruning and causes social behavioral defects. *Mol Psychiatry* 22: 1576–1584.
- Kirsten TB, Casarin RC, Bernardi MM, Felicio LF (2018) Pioglitazone abolishes autistic-like behaviors via the IL-6 pathway. *PLoS One* 13: e0197060.
- Kordulewska NK, Kostyra E, Chwała B, Moszyńska M, Cieślińska A, Fiedorowicz E, Jarmołowska B (2019) A novel concept of immunological and allergy interactions in autism spectrum disorders: Molecular, anti-inflammatory effect of osthole. *Int Immunopharmacol* 72: 1–11.
- Kucukdereli H, Allen NJ, Lee AT, Feng A, Ozlu MI, Conatser LM, et al. (2011) Control of excitatory CNS synaptogenesis by astrocyte-secreted proteins hevin and SPARC. *Proc Natl Acad Sci* 108: E440–449.
- Lammert CR, Lukens JR (2019) Modeling autism-related disorders in mice with maternal immune activation (MIA). *Methods Mol Biol* 1960: 227–236.
- Lee AS, Azmitia EC, Whitaker-Azmitia PM (2017) Developmental microglial priming in postmortem autism spectrum disorder temporal cortex. *Brain Behav Immun* 62: 193–202.
- Lee M, Krishnamurthy J, Susi A, Sullivan C, Gorman GH, Hisle-Gorman E, Erdie-Lalena CR, Nylund CM (2018) Association of autism spectrum disorders and inflammatory bowel disease. *J Autism Develop Disord* 48: 1523–1529.
- Liao TC, Lien YT, Wang S, Huang SL, Chen CY (2016) Comorbidity of atopic disorders with autism spectrum disorder and attention-deficit/hyperactivity disorder. *J Pediatrics*, 171: 248–255.
- Loomes R, Hull L, Mandy WPL (2017) What is the male-to-female ratio in autism spectrum disorder? A systematic review and meta-analysis. *J Am Acad Child Adolesc Psychiatry* 56: 466–474.
- Lucas T, Waisman A, Ranjan R, Roes J, Krieg T, Müller W, et al. (2010) Differential roles of macrophages in diverse phases of skin repair. *J Immunol* 184: 3964–3977.
- Ludvigsson JF, Reichenberg A, Hultman CM, Murray JA (2013) A nationwide study association between celiac disease and the risk of autistic spectrum disorders. *JAMA Psychiatry* 70: 1224–1230.
- Lyll K, Ashwood P, Van de Water J, Hertz-Picciotto I (2014) Maternal immune-mediated conditions, autism spectrum disorders, and developmental delay. *J Autism Develop Disord* 44: 1546–1555.
- Marchezan J, Winkler dos Santos EGA, Deckmann I, dos Santos Riesgo R (2018) Immunological dysfunction in autism spectrum disorder: A potential target for therapy. *Neuroimmunomodulation* 25: 300–319.
- Mantovani A, Sica A, Sozzani S, Allavena P, Vecchi A, Locati M (2004) The chemokine system in diverse forms of macrophage activation and polarization. *Trends Immunol* 25: 677–686.
- Masi A, Quintana DS, Glozier N, Lloyd AR, Hickie IB, Guastella AJ (2015) Cytokine aberrations in autism spectrum disorder: a systematic review and meta-analysis. *Mol Psychiatry* 20: 440–446.
- Matelski L, Van de Water J (2016) Risk factors in autism: Thinking outside the brain. *J Autoimmunity* 67: 1–7.
- Matta SM, Hill-Yardin EL, Crack PJ (2019) The influence of neuroinflammation in autism spectrum disorder. *Brain Behav Immun* 79: 75–90.
- Mills CD (2012) M1 and M2 macrophages: oracles of health and disease. *Crit Rev Immunol* 32: 463–488.
- Molloy CA, Morrow AL, Meinen-Derr J, Dawson G, Bernier R, Dunn M, Hyman SL, McMahon WM, Goudie-Nice J, Hepburn S, et al. (2006) Familial autoimmune thyroid disease as a risk factor for regression in children with autism spectrum disorder: a CPEA Study. *J Autism Dev Disord* 36: 317–324.
- Mordekar SR, Prendergast M, Chattopadhyay AK, Baxter PS (2009) Corticosteroid treatment of behaviour, language, and motor regression in childhood disintegrative disorder. *Eur J Pediatric Neurol* 13: 367–369.
- Morgan JT, Barger N, Amaral DG, Schumann CM (2014) Stereological study of amygdala glial populations in adolescents and adults with autism spectrum disorder. *PLoS One* 9: e110356.
- Morgan JT, Chana G, Pardo CA, Achim C, Semendeferi K, Buckwalter J, Courchesne E, Everall IP (2010) Microglial activation and increased microglial density observed in the dorsolateral prefrontal cortex in autism. *Biol Psychiatry* 68: 368–376.
- Mostafa GA, Shehab AA (2010) The link of C4B null allele to autism and a family history of autoimmunity in Egyptian autistic children. *J Neuroimmunol* 223: 115–119.
- Mouridsen SE, Rich B, Isager T, Jørgen Nedergaard N (2007) Autoimmune diseases in parents of children with infantile autism: A case-control study. *Develop Med Child Neurol* 49: 429–432.
- Murray KN, Edye ME, Manca M, Vernon AC, Oladipo JM, Fasolino V, et al. (2019) Evolution of a maternal immune activation (mIA) model in rats: Early developmental effects. *Brain Behav Immun* 75: 48–59.
- Nadeem A, Ahmad SF, Attia SM, Al-Ayadhi LY, Al-Harbi NO, Bakheet SA (2019) Dysregulated enzymatic antioxidant network in peripheral neutrophils and monocytes in children with autism. *Prog Neuropsychopharmacol Biol Psychiatry* 88: 352–359.
- Nance DM, Sanders VM (2007) Autonomic innervation and regulation of the immune system (1987–2007). *Brain Behav Immun* 21: 736–745.
- Nitschke A, Deonandan R, Konkle ATM (2020) The link between autism spectrum disorder and gut microbiota: A scoping review. *Autism* 24: 1328–1344.
- Orihuela R, McPherson CA, Harry GJ (2016) Microglial M1/M2 polarization and metabolic states. *Br J Pharmacol* 173: 649–665.
- Paglia L (2020) Children diagnosed with “ASD” are first of all... children. *Eur J PediatrDent* 21: 8.
- Pavone L, Fiumara A, Bottaro G, Mazzone D, Coleman M (1997) Autism and celiac disease: failure to validate the hypothesis that a link might exist. *Biol Psychiatry* 42: 72–75.
- Petra AI, Panagiotidou S, Hatzigelaki E, Stewart JM, Conti P, Theoharides TC (2015) Gut-microbiota-brain axis and its effect on neuropsychiatric disorders with suspected immune dysregulation. *Clin Therapeutics* 37: 984–995.
- Purves D, Augustine G, Fitzpatrick D, Katz L, LaMantia A, McNamara J, Williams S (2001) “Neuroscience 2nd edition. Sunderland (ma) Sinauer associates.”

- Rodier PM, Ingram JL, Tisdale B, Nelson S, Romano J (1996) Embryological origin for autism: developmental anomalies of the cranial nerve motor nuclei. *J Comp Neurol* 370: 247–261.
- Román GC, Ghassabian A, Bongers-Schokking JJ, Jaddoe VW, Hofman A, de Rijke YB, Verhulst FC, Tiemeier H (2013) Association of gestational maternal hypothyroxinemia and increased autism risk. *Ann Neurol* 74: 733–742.
- Rosser EC, Mauri C (2015) Regulatory B cells: origin, phenotype, and function. *Immunity* 42: 607–612.
- Russo FB, Freitas BC, Pignatari GC, Fernandes IR, Sebat J, Muotri AR, Beltrão-Braga PCB (2018) Modeling the interplay between neurons and astrocytes in autism using human induced pluripotent stem cells. *Biol Psychiatry* 83: 569–578.
- Saghazadeh A, Ataieina B, Keynejad K, Abdolalizadeh A, Hirbod-Mobarakeh A, Rezaei N (2019a) Anti-inflammatory cytokines in autism spectrum disorders: A systematic review and meta-analysis. *Cytokine* 123: 154740.
- Saghazadeh A, Ataieina B, Keynejad K, Abdolalizadeh A, Hirbod-Mobarakeh A, Rezaei N (2019b) A meta-analysis of pro-inflammatory cytokines in autism spectrum disorders: Effects of age, gender, and latitude. *J Psychiatric Res* 115: 90–102.
- Sakaguchi S, Yamaguchi T, Nomura T, Ono M (2008) Regulatory T cells and immune tolerance. *Cell* 133: 775–787.
- Salam AP, Borsini A, Zunszain PA (2018) Trained innate immunity: a salient factor in the pathogenesis of neuroimmune psychiatric disorders. *Mol Psychiatry* 23: 170–176.
- Saurman V, Margolis KG, Luna RA (2020) Autism spectrum disorder as a brain-gut-microbiome axis disorder. *Dig Dis Sci* 65: 818–828.
- Sciara AN, Beasley B, Crawford JD, Anderson EP, Carrasco T, Zheng S, Ordway GA, Chandley MJ (2020) Neuroinflammatory gene expression alterations in anterior cingulate cortical white and gray matter of males with autism spectrum disorder. *Autism Res* 13: 870–884.
- Serra D, Almeida LM, Dinis TCP (2020) Polyphenols in the management of brain disorders: Modulation of the microbiota-gut-brain axis. *Adv Food Nutr Res* 91: 1–27.
- Shen Y, Li Y, Shi L, Liu M, Wu R, Xia K, Zhang F, Ou J, Zhao J (2020) Autism spectrum disorder and severe social impairment associated with elevated plasma interleukin-8. *Pediatric Res*: 1–7.
- Shin S (2018) Safety of celecoxib versus traditional nonsteroidal anti-inflammatory drugs in older patients with arthritis. *J Pain Res* 11: 3211.
- Singh K, Connors SL, Macklin EA, Smith KD, Fahey JW, Talalay P, Zimmerman AW (2014) Sulforaphane treatment of autism spectrum disorder (ASD). *Proc Natl Acad Sci* 111: 15550–15555.
- Sivamaruthi BS, Suganthi N, Kesika P, Chaiyasut C (2020) The role of microbiome, dietary supplements, and probiotics in autism spectrum disorder. *Int J Environmental Res Public Health* 17: 2647.
- Stanek KR, Youngkin EM, Pyle LL, Raymond JK, Driscoll KA, Majidi S (2019) Prevalence, characteristics, and diabetes management in children with comorbid autism spectrum disorder and type 1 diabetes. *Pediatric Diabetes* 20: 645–51.
- Stefanatos GA, Grover W, Geller E (1995) Case study: corticosteroid treatment of language regression in pervasive developmental disorder. *J Am Acad Child Adolesc Psychiatry* 34: 1107–1111.
- Stevens B, Allen NJ, Vazquez LE, Howell GR, Christopherson KS, Nouri N, KD Micheva, AK Mehalow, AD Huberman, B Stafford (2007) The classical complement cascade mediates CNS synapse elimination. *Cell* 131: 1164–1178.
- Streit WJ (1996) The role of microglia in brain injury. *Neurotoxicology* 17: 671.
- Stutzman D, Dopheide J (2015) Acetylcysteine for treatment of autism spectrum disorder symptoms. *Am J Health-System Pharmacy* 72: 1956–1959.
- Sweeten TL, Bowyer SL, Posey DJ, Halberstadt GM, McDougale CJ (2003) Increased prevalence of familial autoimmunity in probands with pervasive developmental disorders. *Pediatrics* 112: e420–e20.
- Taliou A, Zintzaras E, Lykouras L, Francis K (2013) An open-label pilot study of a formulation containing the anti-inflammatory flavonoid luteolin and its effects on behavior in children with autism spectrum disorders. *Clin Ther* 35: 592–602.
- Tetreault NA, Hakeem AY, Jiang S, Williams BA, Allman E, Wold BJ, Allman JM (2012) Microglia in the cerebral cortex in autism. *J Autism Dev Disord* 42: 2569–2584.
- Theoharides T, Asadi S, Panagiotidou S (2012) A case series of a luteolin formulation (NeuroProtek®) in children with autism spectrum disorders. In: SAGE Publications Sage UK: London, England.
- Tsai SJ, Kuo WW, Liu WH, Yin MC (2010) Antioxidative and anti-inflammatory protection from carnosine in the striatum of MPTP-treated mice. *J Agric Food Chem* 58: 11510–11516.
- Tsilioni I, Theoharides TC (2018) Extracellular vesicles are increased in the serum of children with autism spectrum disorder, contain mitochondrial DNA, and stimulate human microglia to secrete IL-1 β . *J Neuroinflammation* 15: 239.
- Uraz S, Tahan G, Aytakin H, Tahan V (2013) N-acetylcysteine expresses powerful anti-inflammatory and antioxidant activities resulting in complete improvement of acetic acid-induced colitis in rats. *Scand J clinical laboratory investigation* 73: 61–66.
- Williams EC, Zhong X, Mohamed A, Li R, Liu Y, Dong Q, et al. (2014) Mutant astrocytes differentiated from Rett syndrome patients-specific iPSCs have adverse effects on wild-type neurons. *Hum Mol Genet*, 23: 2968–2980.
- Win-Shwe TT, Nway NC, Imai M, Lwin TT, Mar O, Watanabe H (2018) Social behavior, neuroimmune markers and glutamic acid decarboxylase levels in a rat model of valproic acid-induced autism. *J Toxicol Sci* 43: 631–643.
- Vargas DL, Nascimbene C, Krishnan C, Zimmerman AW, Pardo CA (2005) Neuroglial activation and neuroinflammation in the brain of patients with autism. *Ann Neurol* 57: 67–81.
- Volkmar F, Siegel M, Woodbury-Smith M, King B, McCracken J, State M (2014) Practice parameter for the assessment and treatment of children and adolescents with autism spectrum disorder. *J Am Acad Child Adolescent Psychiatry* 53: 237–257.
- Yang Y, Tian J, Yang B (2018) Targeting gut microbiome: A novel and potential therapy for autism. *Life Sciences*, 194: 111–119.
- Yizhar O, Fenno LE, Prigge M, Schneider F, Davidson TJ, O'Shea DJ, Sohal VS, Goshen I, Finkelstein J, Paz JT (2011) Neocortical excitation/inhibition balance in information processing and social dysfunction. *Nature*, 477: 171–78.
- Zhang ZR, Leung WN, Cheung HY, Chan CW (2015) Osthole: a review on its bioactivities, pharmacological properties, and potential as alternative medicine. *Evid Based Complement Alternat Med* 2015: 919616.
- Zhang M, Hutter G, Kahn SA, Azad TD, Gholamin S, Xu CY, et al. (2016) Anti-CD47 treatment stimulates phagocytosis of glioblastoma by M1 and M2 polarized macrophages and promotes M1 polarized macrophages in vivo. *PLoS One* 11: e0153550.
- Zhao H, Wang Q, Yan T, Zhang Y, Xu HJ, Yu HP, et al. (2019) Maternal valproic acid exposure leads to neurogenesis defects and autism-like behaviors in non-human primates. *Transl Psychiatry* 9: 267.